

Plant nitrogen acquisition and interactions under elevated carbon dioxide: impact of endophytes and mycorrhizae

XIN CHEN^{*†1}, CONG TU^{†1}, MICHAEL G. BURTON[‡], DOROTHY M. WATSON[‡], KENT O. BURKEY^{‡§} and SHUIJIN HU[†]

^{*}College of Life Sciences, Zhejiang University, Hangzhou 310029, China, [†]Department of Plant Pathology, North Carolina State University, Raleigh, NC 27695, USA, [‡]Department of Crop Science, North Carolina State University, Raleigh, NC 27695, USA, [§]USDA-ARS Plant Science Research Unit, 3127 Ligon Street, Raleigh, NC 27607, USA

Abstract

Both endophytic and mycorrhizal fungi interact with plants to form symbiosis in which the fungal partners rely on, and sometimes compete for, carbon (C) sources from their hosts. Changes in photosynthesis in host plants caused by atmospheric carbon dioxide (CO₂) enrichment may, therefore, influence those mutualistic interactions, potentially modifying plant nutrient acquisition and interactions with other coexisting plant species. However, few studies have so far examined the interactive controls of endophytes and mycorrhizae over plant responses to atmospheric CO₂ enrichment. Using *Festuca arundinacea* Schreb and *Plantago lanceolata* L. as model plants, we examined the effects of elevated CO₂ on mycorrhizae and endophyte (*Neotyphodium coenophialum*) and plant nitrogen (N) acquisition in two microcosm experiments, and determined whether and how mycorrhizae and endophytes mediate interactions between their host plant species. Endophyte-free and endophyte-infected *F. arundinacea* varieties, *P. lanceolata* L., and their combination with or without mycorrhizal inocula were grown under ambient (400 µmol mol⁻¹) and elevated CO₂ (ambient + 330 µmol mol⁻¹). A ¹⁵N isotope tracer was used to quantify the mycorrhiza-mediated plant acquisition of N from soil. Elevated CO₂ stimulated the growth of *P. lanceolata* greater than *F. arundinacea*, increasing the shoot biomass ratio of *P. lanceolata* to *F. arundinacea* in all the mixtures. Elevated CO₂ also increased mycorrhizal root colonization of *P. lanceolata*, but had no impact on that of *F. arundinacea*. Mycorrhizae increased the shoot biomass ratio of *P. lanceolata* to *F. arundinacea* under elevated CO₂. In the absence of endophytes, both elevated CO₂ and mycorrhizae enhanced ¹⁵N and total N uptake of *P. lanceolata* but had either no or even negative effects on N acquisition of *F. arundinacea*, altering N distribution between these two species in the mixture. The presence of endophytes in *F. arundinacea*, however, reduced the CO₂ effect on N acquisition in *P. lanceolata*, although it did not affect growth responses of their host plants to elevated CO₂. These results suggest that mycorrhizal fungi and endophytes might interactively affect the responses of their host plants and their coexisting species to elevated CO₂.

Keywords: elevated CO₂, endophyte, *Festuca arundinacea*, mycorrhizae, ¹⁵N tracer, plant N acquisition, *Plantago lanceolata*

Received 6 March 2006; revised version received 20 September 2006 and accepted 28 November 2006

Correspondence: Shuijin Hu, fax +1 919 513 1279, e-mail: Shuijin_hu@ncsu.edu

¹These authors contributed equally to this work.

Introduction

Carbon dioxide (CO₂) enrichment in the atmosphere in general stimulates photosynthetic activity and growth of C₃ plants. This may in turn alter the availability of photosynthates for plant-associated microbes (Hu *et al.*, 1999), modifying plant–microbial interactions such as

plant–rhizobial symbiosis (Zanetti *et al.*, 1996), mycorrhizae (Johnson *et al.*, 2003), and plant–endophyte complexes (Newman *et al.*, 2003; Hunt *et al.*, 2005).

More than 80% of the higher plant species examined are associated with arbuscular mycorrhizal (AM) fungi (Smith & Read, 1997). Many studies have shown that elevated CO₂ often increases mycorrhizal colonization of roots and to a lesser degree mycorrhizal activities (Rillig *et al.*, 1999; Hartwig *et al.*, 2002; Treseder, 2004; Hu *et al.*, 2005). This CO₂-enhancement may alter plant nutrient uptake and plant interactions with their neighbors (van der Heijden *et al.*, 1998; Smith *et al.*, 1999; O'Connor *et al.*, 2002), particularly if mycorrhizae in the coexisting species have differential responses to elevated CO₂.

Similar to mycorrhizal fungi, endophytic fungi within asymptomatic aerial tissues of plants represent a ubiquitous component of terrestrial plant communities (Arnold *et al.*, 2003). About two-thirds of cold-season grasses such as tall fescue (*Festuca arundinacea* Schreb) and ryegrass (*Lolium perenne*) form symbionts in their aboveground parts with endophytic fungi (Müller & Krauss, 2005). For example, tall fescue is often associated with a systemic fungal endophyte, *Neotyphodium coenophialum* Morgan-Jones and Gams (Ascomycota: Clavicipitaceae) (Bacon *et al.*, 1977; Glenn *et al.*, 1996). Endophytes often enhance the resistance of the host plants to environmental stresses such as drought (Richardson *et al.*, 1993; Schardl *et al.*, 2004) and pathogens (Arnold *et al.*, 2003). Endophytic fungi in tall fescue produce a group of toxic compounds named ergot alkaloids (Hill *et al.*, 1994) that protect host plants from herbivores (Bree, 1994; Clay & Schardl, 2002), fungi (Latch, 1993) and nematodes (Gwinn & Bernard, 1993) and increase host resistance to both water and nitrogen (N) stress (Lyons *et al.*, 1990; Elmi & West, 1995; Clay & Schardl, 2002), resulting in increased host tolerance to marginal soil environments.

Both mycorrhizal and endophytic fungi solely rely on C sources from their host plants. In some plants, endophytic and mycorrhizal fungi may compete for photosynthates (Vicari *et al.*, 2002). Therefore, factors that alter plant photosynthesis and subsequent allocation of photosynthetic products may profoundly influence the colonization of those mutualistic fungi, as well as their functioning. Because the CO₂ concentration is a limiting factor for many C₃ plants, the rising CO₂ concentration in the atmosphere can enhance the availability of photosynthates for both mycorrhizal (Staddon & Fitter, 1998; Johnson *et al.*, 2003; Treseder, 2004) and endophytic fungi (Marks & Clay, 1990). Over the last decade, many experiments have examined the effect of elevated CO₂ on mycorrhizae (see recent meta-analysis by Treseder, 2004) and the subsequent effects on host

growth. However, only a few experiments have explicitly investigated the role of endophytic fungi in mediating the response of their host grasses to elevated CO₂ (Marks & Clay, 1990; Marks & Lincoln, 1996; Applebee *et al.*, 1999; Newman *et al.*, 2003; Hunt *et al.*, 2005). Among over 50 publications on perennial ryegrass, only two examined the status of endophytic fungi (Marks & Lincoln, 1996; Hunt *et al.*, 2005). In tall fescue, Newman *et al.* (2003) examined the response of endophyte-infected (E+) and endophyte-free (E−) plants to elevated CO₂ and found that the presence of endophytes modify plant response by altering N metabolism. In spite of common existence of both mycorrhizae and endophytes in cold-season grasses, to our knowledge, no experiments have explicitly examined any potential impacts of elevated CO₂ on the interaction between endophytes and mycorrhizae.

In the present study, we examined how elevated CO₂ affects endophytes and mycorrhizae, and their functions, using tall fescue and *Plantago lanceolata* as model species. Tall fescue is one of the most important perennial grasses, occupying more than 35 million ha as a forage, turf or soil conservation plant in the United States alone (Ball *et al.*, 1993; Rudgers *et al.*, 2004). Because of high tolerance to environmental stresses and adaptation to various climatic conditions, tall fescue is a common species in many natural ecosystems and is an invasive species to some endangered habitats across the world (Clay & Holah, 1999; Saikkonen, 2000; Rudgers *et al.*, 2004). In general, tall fescue is not readily colonized by mycorrhizae and the cause for low mycorrhizal infection is not exactly known. It was suggested that endophytes compete with mycorrhizae for carbon resources within the host or that endophytes are inhibiting AM colonization through fungitoxic compounds (Chu-Chou *et al.*, 1992; Guo *et al.*, 1992; Matthews & Clay, 2001; Müller, 2003). A few recent studies also showed that *F. arundinacea* is very responsive to elevated CO₂ (Newman *et al.*, 2003), but whether CO₂-enhancement of C availability will alter mycorrhizal infection and activities in fescue roots remains unexplored. Tall fescue coexists with many cool-season grasses and forbs in pastures and natural habitats. *Plantago* (*P. lanceolata* L.) is a common weedy species in many pastures and natural habitats dominated by tall fescue across the world (Barton & Nestor, 1979). It is highly mycorrhizal (Staddon *et al.*, 1999; Hodge *et al.*, 2001) and is very responsive to CO₂ enrichment (Rouhier & Read, 1998). An understanding of CO₂ effects on mycorrhizae and endophytes will provide insight into the role of those symbiotic fungi on resource acquisition/partitioning and interactions among coexisting host plant species under anticipated CO₂ scenarios.

We conducted two experiments investigating how mycorrhizae and endophytes mediate the response of tall fescue and *P. lanceolata* to elevated CO₂ both in monoculture and mixture. The specific objectives of the experiment were to examine (1) whether endophytes affect mycorrhizal responses to elevated CO₂ in *F. arundinacea* and *P. lanceolata*, and (2) whether and how endophytes and mycorrhizae mediate nutrient acquisition and interactions of their host plants under elevated CO₂.

Materials and methods

The CO₂ facility

A microcosm study was conducted in the CO₂ exposure facility at the USDA Air-Quality greenhouse at North Carolina State University. The CO₂ facility consists of a 9 m × 12 m greenhouse bay containing 20 continuously stirred tank reactor (CSTR) chambers each measuring 1.2 m diameter × 1.4 m tall for exposure of plants to CO₂ gas (Booker *et al.*, 2000). The chambers were made of transparent materials that allowed full light penetration. Dispensing and monitoring of gas treatments are accomplished in a laboratory adjacent to the greenhouse. A blower system provided a constant flow of charcoal filtered air through each CSTR. For those chambers assigned to an elevated CO₂ treatment, compressed CO₂ was added to the air entering the CSTR using a rotameter to control flow so that CO₂ concentration was maintained at target level. The air continuously moved out the CSTR and thus alleviated the heating effect of chambers. Monitoring of CO₂ concentration was accomplished using computer-activated solenoid valves to direct gas exiting the CSTR into infrared analyzers (model 6252, LiCor Inc., Lincoln, NE, USA). The computer collected and averaged temperature and CO₂ data for analysis.

Experiment 1: effects of mycorrhizae on F. arundinacea and P. lanceolata responses to elevated CO₂ in monoculture and mixture

Plant seeds and mycorrhizal fungal inocula. Seeds of endophyte-free tall fescue (cv. Kentucky-31) (E–K) and *P. lanceolata* (PL) were obtained from commercial suppliers (The Wyatt-Quarles Seed Co., Raleigh, NC, USA, and The Nurseries, Billingbear Park, Wokingham, Berkshire, UK, respectively). Mycorrhizal inoculum was obtained from a field at the Center for Environmental Farming Systems, North Carolina State University, near Goldsboro, NC, USA. The AM inoculum was a mixture of soil with spores, hyphae and root residues containing *Glomus etunicatum*, *Glomus*

clarum, *Gigaspora margarita*, *Gigaspora rosea*, *Scutellospora heterogama*, *Scutellospora pellucida*, *Acaulospora scrobiculata*, *Acaulospora laevis*, *Acaulospora koskei*, *Acaulospora mellea* and *Acaulospora* sp.

Experimental design and treatments. The experiment was established in a split-split plot design with atmospheric CO₂ concentrations (ambient and elevated CO₂) as the mainplot factor, mycorrhizae (mycorrhizae and non-mycorrhizae) as subplot factors, and plants (E–K, PL and their mixture) as sub-subplot factors. Six CSTR chambers were blocked into three blocks (i.e. three chambers per CO₂ level). Either an ambient (400 µmol mol⁻¹) or elevated CO₂ level (ambient + 330 µmol mol⁻¹) was randomly assigned to each chamber within a block. The elevated CO₂ concentration corresponds approximately to the predicted CO₂ concentration by the end of this century (IPCC, 2001). Two microcosm units (one with mycorrhizae and another with no mycorrhizae) were placed in each chamber.

A microcosm unit containing six 2.7 L compartments (described in detail by Tu *et al.*, 2006) was employed with three compartments on one side designated as Host and the other three as Test compartments. Each Host/Test pair in a microcosm was separated by a replaceable mesh that allowed penetration by hyphae but not roots. The soil used was a mixture of one part of a sandy loam soil with two parts of pure sand (by weight). The mixture had an extractable N of 7.6 mg N kg⁻¹ and a pH of 6.7. Each compartment of the microcosm was filled with 3.0 kg of the sterilized soil mixture. For mycorrhizal microcosms, 10.0 g AM inoculum was incorporated into the soil in each Host compartment. The nonmycorrhizal microcosms received 100 mL of microbial filtrate solution filtrated by Whatman No. 1 filter paper from 10.0 g AM inoculum (with no mycorrhizal spores) and the sterilized residues of the inoculum to correct for possible differences in the microbial community and nutrients between mycorrhizal and nonmycorrhizal treatments.

Seeds of *P. lanceolata* (20 seeds), *F. arundinacea* (E–K, 40 seeds) and their mixture (10 and 20 seeds, respectively) were respectively sown in the three Host compartments in each microcosm. After emergence, E–K and PL seedlings were thinned to ensure the same number of plants in each compartment. The plants were grown in ambient light and temperature conditions. The plants were watered with distilled water daily. The TEST compartments were also watered but less frequently in order to maintain the soil moisture so that AM fungal hyphae could grow into the Test soils. Eight weeks after seeding, mineral N (NH₄NO₃) was added at a rate of 10 mg N kg⁻¹ soil to each Host and Test compartment.

¹⁵N tracer. To quantify mycorrhizally mediated plant N uptake, the ¹⁵N tracer was injected uniformly as ¹⁵N-enriched mineral N ((NH₄)₂SO₄, 99.7% atom ¹⁵N) in DI water at a rate of 3.0 mg N kg⁻¹ soil into each Test compartment 1 week before harvesting.

Collection and analyses of plant and soil samples. The plants were harvested 14 weeks after seeding. Root systems were separated from shoots and the fresh root biomass was weighed immediately. Roots of the two species in the mixture were also visually separated. Half of each fresh root sample was stored at 4 °C for quantification of AM fungal colonization and the remaining half was oven dried (65 °C for 48 h) and used for biomass measurement. Soil samples in the Host and Test compartments were collected separately and stored at 4 °C for extractable N quantification.

The fresh shoot biomass was weighed immediately following the harvest. Ten tillers from each sample were stored at -20 °C until they were tested for the presence of endophytes. The remaining shoots were dried at 65 °C for 48 h and weighed, and the total shoot biomass was obtained by adding the dry weight of those 10 tillers later. The oven-dried shoots were used for measurements of total N and ¹⁵N. Root biomass C was based on ash-free dry weight because of potential soil contamination. The C concentration of roots was analyzed on a Perkin-Elmer 2400 CHNS/O elemental analyzer (Norwalk, CT, USA). Root biomass C was calculated by multiplying the C concentration of roots and the total weight of roots.

Experiment 2: Endophytic effects on responses of F. arundinacea and P. lanceolata to elevated CO₂ in mixture

Seeds and mycorrhizal fungal inocula. Seeds of *P. lanceolata* were from the same seedlot as was used in Experiment 1. Three varieties of *F. arundinacea* [e.g. endophyte-free cv. Jesup (E-J), endophyte-infected cv. Jesup (E + J) and endophyte-infected cv. Kentucky-31 (E + K)], were used to examine endophyte effects on responses to elevated CO₂ of both *P. lanceolata* and *F. arundinacea* growing in mixture. The E-J and E + J seeds were provided by Pennington Seed Inc. (Madison, GA, USA), and the E + K seeds were purchased from the Wyatt-Quarles Seed Co. While cultivar E-J was the endophyte-free isogenic line of cultivar (E + J), E + K was the same variety of E-K in Experiment 1 except for the endophyte status. Mycorrhizal inoculum was a mixture of soil with spores, hyphae and root residues containing *Glomus intraradices*, *G. margarita* and *S. heterogama*, which had the same original source as that in the Experiment 1.

Experimental design and treatments. The experiment was a split-plot design with atmospheric CO₂ concentrations (ambient and elevated CO₂) as the mainplot factor, and endophyte/variety (E-J, E + J and E + K) as subplot factors. Either ambient (400 µmol mol⁻¹) or elevated CO₂ level (ambient + 330 µmol mol⁻¹) was randomly assigned to four pairs of chambers (i.e. four replicates per CO₂ level). Each compartment of the microcosm units was filled with 3.0 kg of the sterilized soil mixture and then the Host compartment inoculated with 10.0 g of mycorrhizal inocula. Two microcosm units were placed in each chamber to provide two levels of additional N inputs (10 and 35 mg N kg⁻¹ soil) in which mineral N (NH₄NO₃) in solution was added to both Test and Host compartments.

Seeds of *P. lanceolata* (10 seeds) and *F. arundinacea* (20 seeds) of E-J, E + J or E + K were sown into each Host compartment. Seedlings of *P. lanceolata* and *F. arundinacea* plants in each compartment were then thinned to ensure each compartment have exactly same numbers of plants. The plants were managed similarly as in Experiment 1 and this experiment was terminated 18 weeks after seeding. Plant samples were collected for biomass and analyses.

¹⁵N tracer. The ¹⁵N at a rate of 3.0 mg N kg⁻¹ soil in form of (NH₄)₂SO₄ (99.7% atom ¹⁵N) solution was injected uniformly in each Test compartment 3 weeks before harvesting to quantify mycorrhizally mediated plant N uptake.

Measurements of ¹⁵N and N in soil and plant biomass

A subsample (ca. 1.0 g) of each dried shoot sample was ground using an 8000D SPEX CertiPrep Dual Mixer (SPEX CertiPrep, Metuchen, NJ, USA). Shoot N concentrations and N isotope ratios (¹⁴N and ¹⁵N) were determined using a ThermoFinnigan DELTAPlus continuous flow isotope ratio mass spectrometer (CF-IRMS, Thermo Finnigan DELTA Plus, Waltham, MA, USA). Sample ¹⁵N concentrations were calculated from fractional abundance [¹⁵N/(¹⁴N + ¹⁵N)] and total N content (Hu *et al.*, 2001). The amount of plant ¹⁵N uptake mediated by mycorrhizae was calculated by ¹⁵N in mycorrhizal samples minus ¹⁵N in nonmycorrhizal samples. Ammonium (NH₄⁺) and nitrate (NO₃⁻) in the soil were extracted using 0.5 M K₂SO₄ and their concentrations were determined using a fluid injection autoanalyzer (Lachat Instruments, Milwaukee, WI, USA).

Mycorrhizal colonization of plant roots

Root colonization by mycorrhizal fungi was quantified using a dissection microscope (× 20–40) after cleaning

the roots in 10% KOH (w/v) and staining with trypan blue in an acid glycerol solution. A variation of the gridline intersection method, developed by Giovannetti & Mosse (1980), was used to determine the percentage of root length in which arbuscles, vesicles or hyphae occurred.

Presence of endophytes in tall fescue seeds and plants

The presence of endophyte in both tillers and seeds was determined by a histological staining method (Hiatt *et al.*, 1999). Before the seeds were planted, 100 tall fescue seeds were soaked in a 5% NaOH solution at room temperature for 12–16 h. The seeds were rinsed thoroughly in running tap water and were then stained with aniline blue-lactic acid solution. The stained seeds were crushed slightly on slides and examined under a compound microscope. Plants collected at the final harvest were further examined for the existence of endophytes. The longitudinal sections from each frozen tiller sheath were stained with the aniline blue-lactic acid solution and then examined using a compound microscope.

Statistical analysis

Data from each experiment were subjected to analysis of variance using the general linear model procedure of the STATISTICAL ANALYSIS SYSTEM package (v. 8.02, SAS Institute, Cary, NC, USA) with appropriate error term to either a split-split plot (Experiment 1) or a split plot (Experiment 2) design. The significant differences were determined at the 95% probability level.

Results

*Experiment 1: effects of mycorrhizae on responses of *F. arundinacea* and *P. lanceolata* to elevated CO₂ in monoculture and mixture*

*Shoot biomass, root biomass and shoot biomass ratio (PL : FA) of *P. lanceolata* and *F. arundinacea*.* Elevated CO₂ significantly increased shoot biomass, but mycorrhizae did not influence shoot biomass (Fig. 1a, $P > 0.68$). In monoculture, elevated CO₂ enhanced shoot biomass of both *F. arundinacea* and *P. lanceolata* with larger effects on *P. lanceolata* (with a 52.0% increase) than on *F. arundinacea* (with a 30.4% increase) (Fig. 1a), regardless of the mycorrhizal status. In the mixture, total shoot biomass increased significantly under elevated CO₂ (Fig. 1a), and no significant mycorrhizal effect was observed (Fig. 1a). Both elevated CO₂ and mycorrhizae significantly increased the PL : FA ratio of shoot biomass in the mixture (Fig. 2a). Also, root

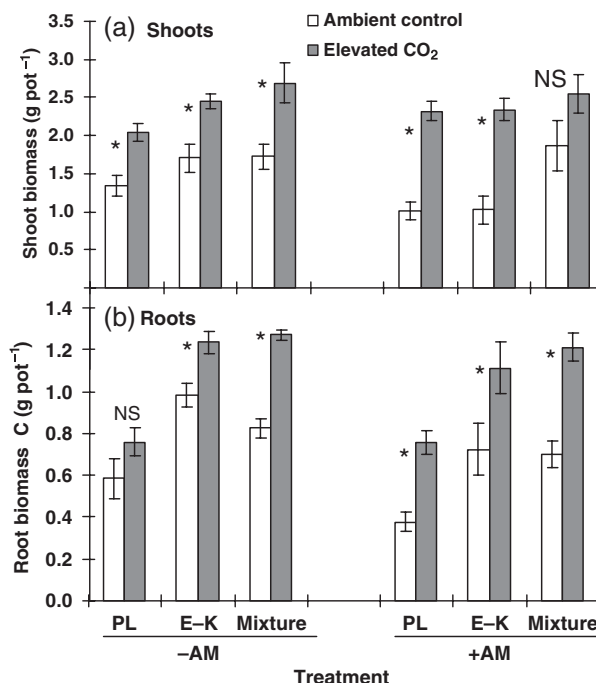


Fig. 1 Plant biomass as influenced by elevated carbon dioxide (CO₂) and arbuscular mycorrhizae (AM). (a) Shoot biomass and (b) root biomass in Experiment 1. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$; NS, not significant between ambient and elevated CO₂; PL, *Plantago lanceolata* alone; E-K, *F. arundinacea*, endophyte-free cv. Kentucky-31 alone; Mixture, PL and E-K together.

biomass C was significantly increased by elevated CO₂ ($P < 0.02$) (Fig. 1b).

Shoot biomass N and biomass ¹⁵N. Elevated CO₂ did not significantly affect total shoot biomass N ($P > 0.53$), but AM fungi and plant species significantly affected shoot biomass N ($P < 0.05$ and $P < 0.001$, respectively; Fig. 3). Mycorrhizae significantly enhanced shoot biomass N of *P. lanceolata*, altering N distribution between *P. lanceolata* and *F. arundinacea* in the mixture. Under the ambient CO₂, biomass N in *P. lanceolata* shoots only accounted for 13.7% and 17.5% of total biomass N in the absence and presence of mycorrhizae, respectively. However, under elevated CO₂, this number increased to 26.9% and 40.3%, respectively (Fig. 3).

Mycorrhizae significantly increased ¹⁵N uptake of *P. lanceolata* but not of *F. arundinacea* under both CO₂ concentrations. This mycorrhizal effect was also significantly affected by CO₂ concentrations (Fig. 4). Compared with the ambient CO₂, elevated CO₂ significantly increased ¹⁵N uptake of *P. lanceolata* both in the monoculture and in the mixture, but had no effect on shoot ¹⁵N in *F. arundinacea* (Fig. 4). Because *P. lanceolata* only consisted of a small proportion of the

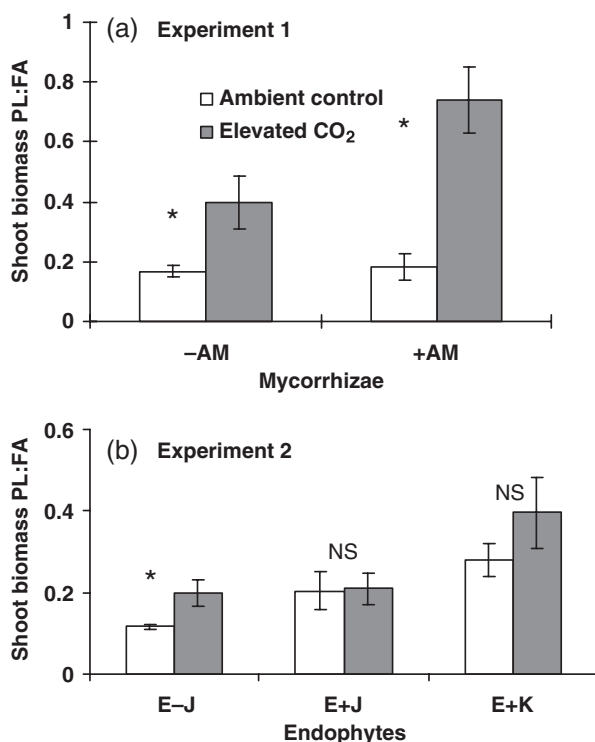


Fig. 2 Effects of arbuscular mycorrhizae (AM) (a) or endophytes (b) on the PL:FA ratio of shoot biomass in mixture under elevated carbon dioxide (CO₂). PL, *Plantago lanceolata*; FA, *Festuca arundinacea*; E-J, endophyte-free cv. Jesup; E+J, endophyte-infected cv. Jesup; E+K, endophyte-infected cv. Kentucky-31. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$ between ambient and elevated CO₂.

total biomass, mycorrhizal enhancement of total shoot ¹⁵N content was reduced in the mixture (Fig. 4).

Soil extractable N in Host and Test compartments. No significant differences in soil extractable N in the Host soils were found among all treatments (data not shown), with the average of soil extractable N being at about 2.0 mg N kg⁻¹ soil. However, soil extractable N in the Test soils was reduced by mycorrhizae, with significant reduction only in soils with *F. arundinacea* under ambient CO₂ and with *P. lanceolata* under elevated CO₂ ($P < 0.05$, Fig. 5). Compared with their respective ambient controls, elevated CO₂ tended to reduce extractable N in the Test soils in all treatments (Fig. 5), but this effect was not statistically significant ($P = 0.21$).

Mycorrhizal colonization of plant roots and the presence of endophyte in tillers and seeds. No obvious mycorrhizal colonization was found in the roots of both plant species in the no-mycorrhizal treatments under either CO₂ levels. When inoculated with mycorrhizae,

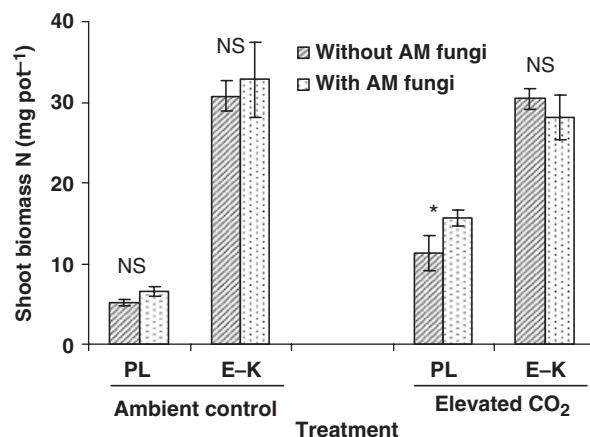


Fig. 3 Elevated carbon dioxide (CO₂) and arbuscular mycorrhiza effects on nitrogen (N) distribution between *Plantago lanceolata* (PL) and *Festuca arundinacea* (E-K) in their mixture in the Experiment 1. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$.

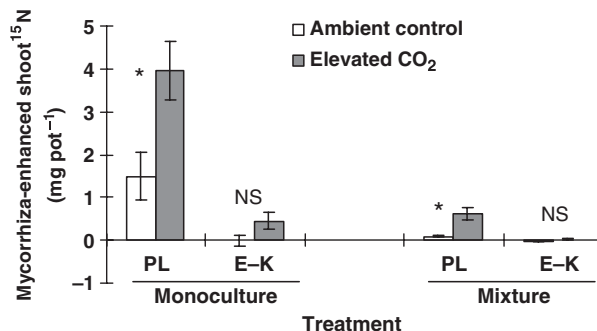


Fig. 4 Mycorrhiza-mediated ¹⁵N acquisition of *Plantago lanceolata* and *Festuca arundinacea* in monoculture or mixture under ambient and elevated carbon dioxide (CO₂) in the Experiment 1. Shoot ¹⁵N values refer to the ¹⁵N excess in shoot biomass compared with the corresponding no-mycorrhizae control and were obtained by subtracting shoot ¹⁵N content in the no-mycorrhizae control from the mycorrhizal treatment. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$; NS, not significant between ambient and elevated CO₂. Abbreviations refer to Fig. 1.

mycorrhizal root colonization of *F. arundinacea* was still very low (<5%) both in monoculture and in mixture, regardless of atmospheric CO₂ concentrations (data not shown). In contrast, mycorrhizal root colonization of *P. lanceolata* was 31.0% in monoculture and 27.0% in mixture under ambient CO₂, and, respectively, increased to 46.0% and 37.0% under elevated CO₂ (Fig. 6a). Total root length colonized by mycorrhizal fungi tended to increase under elevated CO₂ but this effect was not statistically significant ($P = 0.24$). As to the endophytic status of *F. arundinacea*, neither the seeds nor the tillers collected at the end of the experiment were infected with endophytes.

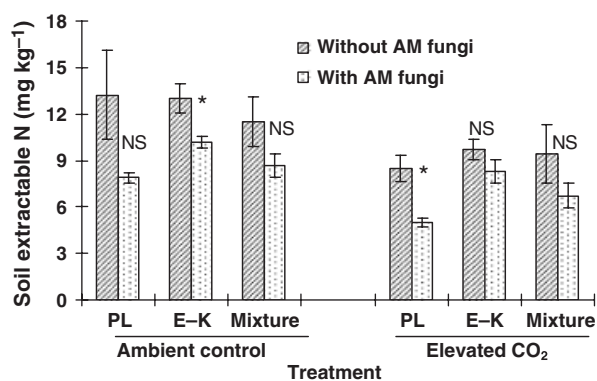


Fig. 5 Soil extractable nitrogen (N) in the Test compartments as affected by elevated CO₂ and arbuscular mycorrhizae (AM) in Experiment 1. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$; NS, not significant between mycorrhizae (+AM) and no-mycorrhizae (-AM). Abbreviations refer to Fig. 1.

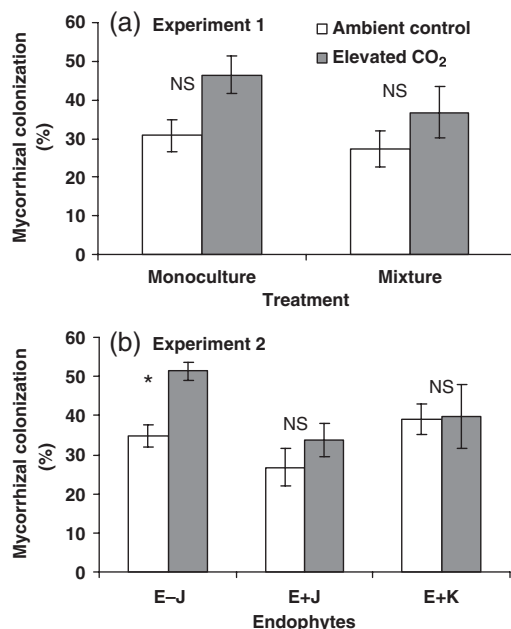


Fig. 6 Effects of elevated CO₂ on mycorrhizal root colonization (MRC) of *Plantago lanceolata* both in monoculture and in mixture with different fescue varieties. (a) Monoculture and mixture with endophyte-free fescue plants in Experiment 1. (b) Mixture with fescue varieties of different endophytic status in Experiment 2. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$; NS, not significant between ambient and elevated CO₂.

Experiment 2: endophytic effects on responses of *F. arundinacea* and *P. lanceolata* to elevated CO₂ in plant mixtures

Shoot biomass, root biomass and shoot biomass ratio (PL : FA) of *P. lanceolata* and *F. arundinacea* in the mixture. Endophytes influenced shoot biomass of *P. lanceolata*

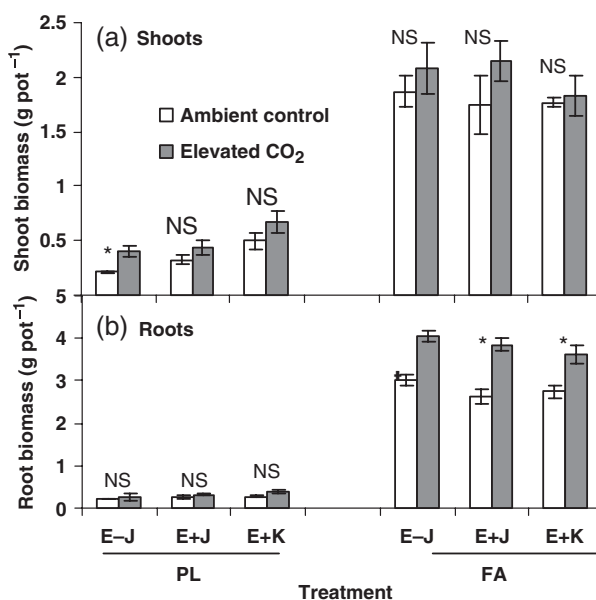


Fig. 7 Effects of fescue variety/endophytic status on shoot (a) and root (b) biomass of *Plantago lanceolata* (PL) and *Festuca arundinacea* (FA) in the mixture in Experiment 2. E-J, endophyte-free cv. Jesup; E+J, endophyte-infected cv. Jesup; E+K, endophyte-infected cv. Kentucky-31. Bars depict treatment means and standard errors (SE). * $P \leq 0.05$; NS, not significant between ambient and elevated CO₂.

($P < 0.01$), but not *F. arundinacea* ($P = 0.9$) (Fig. 7a). Compared with the E-control (PL ~ E-J), shoot biomass of *P. lanceolata* under both ambient and elevated CO₂ was significantly higher in the PL ~ E+J and PL ~ E+K mixtures with an average increase of 23% and 90%, respectively. This led to a significant increase in the shoot biomass ratios of PL : FA in the presence of endophytes (Fig. 2b, $P < 0.003$). Elevated CO₂ significantly increased shoot biomass of *P. lanceolata* in the PL ~ E-J combination but had no significant effects on *F. arundinacea* shoot biomass (Fig. 7a), increasing the PL : FA biomass ratios in the PL ~ E-J combination (Fig. 2b). However, this CO₂ effect was offset by endophytes as the PL : FA ratios were not affected by CO₂ enrichment in the PL ~ E+J and PL ~ E+K mixtures (Fig. 2b). Also, CO₂ enrichment enhanced root biomass of *F. arundinacea* but not *P. lanceolata* (Fig. 7b). The presence of endophytes reduced root biomass of host fescue plants by 8–10% ($P = 0.7$), but increased root biomass of *P. lanceolata* by 18–38% ($P < 0.02$) across the CO₂ levels (Fig. 7b).

Shoot biomass N and ¹⁵N. Elevated CO₂ significantly reduced shoot N concentrations of both *P. lanceolata* and *F. arundinacea* (Table 1), but only led to a significant decrease in total shoot biomass N of

Table 1 Effects of elevated carbon dioxide (CO₂) on shoot ¹⁵N and shoot N concentrations of *Plantago lanceolata* and *Festuca arundinacea* grown in mixture in Experiment 2

Variety	¹⁵ N concentration (μg ¹⁵ N kg ⁻¹)		N concentration (mg N kg ⁻¹)	
	ACO ₂	ECO ₂	ACO ₂	ECO ₂
<i>P. lanceolata</i>				
E-J	53.9 ± 3.31	71.0 ± 11.6	10.9 ± 0.36	10.8 ± 1.70
E+J	49.9 ± 1.25	43.5 ± 4.35	10.2 ± 0.56	8.26 ± 0.36*
E+K	59.5 ± 5.33	49.0 ± 7.64	11.7 ± 0.74	8.98 ± 1.35*
<i>F. arundinacea</i>				
E-J	44.4 ± 1.96	38.0 ± 2.92	10.4 ± 0.36	8.45 ± 0.44*
E+J	48.5 ± 0.51	38.8 ± 3.03	10.9 ± 0.31	8.30 ± 0.27*
E+K	56.6 ± 8.44	47.2 ± 6.43	11.4 ± 0.48	8.53 ± 0.33*

*Significantly different from the corresponding ambient control according to LSD test ($P \leq 0.05$).

ACO₂, ambient CO₂; ECO₂, elevated CO₂; N, nitrogen.

Table 2 Effects of elevated carbon dioxide (CO₂) on ¹⁵N and N accumulation in the shoot biomass of *Plantago lanceolata* and *Festuca arundinacea* in mixture in Experiment 2

Variety	Biomass ¹⁵ N (μg N pot ⁻¹)			Biomass N (mg N pot ⁻¹)		
	ACO ₂	ECO ₂	Means [†]	ACO ₂	ECO ₂	Means
<i>P. lanceolata</i>						
E-J	11.5 ± 0.35	28.5 ± 5.33*	20.0b	2.35 ± 0.12	4.25 ± 0.66*	3.30b
E+J	15.9 ± 1.94	19.3 ± 3.97	17.6b	3.28 ± 0.49	3.60 ± 0.56	3.44b
E+K	29.1 ± 4.82	33.5 ± 7.80	31.3a	5.81 ± 1.01	6.22 ± 1.47	6.01a
<i>F. arundinacea</i>						
E-J	82.2 ± 3.53	77.2 ± 2.96	79.7a	19.3 ± 1.29	17.3 ± 1.28	18.3a
E+J	84.9 ± 12.9	82.4 ± 6.30	83.6a	19.2 ± 3.12	17.8 ± 1.36	18.5a
E+K	100.4 ± 16.5	86.0 ± 13.9	93.2a	20.1 ± 0.95	15.6 ± 1.78*	17.9a

[†]Means are averages of shoot biomass N in each fescue variety treatment under both ambient and elevated CO₂; Values followed by different letters within a plant are significantly different among the fescue varieties ($P \leq 0.05$).

*Significantly different from the corresponding ambient control within a measurement (LSD, $P \leq 0.05$).

ACO₂, Ambient CO₂; ECO₂, Elevated CO₂; N, nitrogen.

F. arundinacea in the PL~E+K mixture (Table 2). CO₂ enrichment significantly increased total biomass N and biomass ¹⁵N in *P. lanceolata* shoots in the absence of endophytes (i.e. E-J), but not in the presence of endophytes (i.e. E+J and E+K) (Table 2). The presence of endophytes in fescue plants did not influence shoot N concentrations of either plant species, but increased total shoot N of *P. lanceolata* with statistical significance in the E+K treatment (Table 2). No significant effects of endophytes were observed on the ¹⁵N concentrations or total shoot ¹⁵N of either plant species (by the E-J vs. E+J comparison) (Table 2).

Mycorrhizal colonization of plant roots and the presence of endophyte in tillers and seeds. Similar to what observed in the Experiment 1, mycorrhizal colonization of fescue roots was again very low (<3%) and was not affected by

CO₂ levels and fescue varieties (data not shown). Likewise, fescue varieties (i.e. endophytic status) had no effects on the mycorrhizal root colonization and total colonization length of *P. lanceolata* under either CO₂ concentrations. However, elevated CO₂ enhanced the AM root colonization of *P. lanceolata* only when they were grown together with endophyte-free fescue plants (Fig. 6a and b). In addition, endophytic fungi were detected in all fescue plants in both E+J and E+K cultivars, but not in the E-J plants.

Effects of N availability on plant biomass, total plant N and plant ¹⁵N content. High N input (35 mg N kg⁻¹ soil) significantly ($P < 0.05$) increased the biomass, N concentrations and total N contents, and ¹⁵N contents of both *P. lanceolata* and *F. arundinacea* (data not shown).

However, no significant interactions were observed between N availability and endophytes.

Discussion

Results from our experiments showed that elevated CO₂ stimulated the growth of both plantago and tall fescue plants (Figs 1 and 7). However, the magnitude of the stimulation was significantly higher in plantago than in tall fescue, altering the relative composition of species biomass in mixture (Fig. 2). These results are consistent with a general pattern observed in other studies in which CO₂ enrichment favors forbs over grasses (Potvin & Vasseur, 1997; Leadley *et al.*, 1999; Owensby *et al.*, 1999; Teyssonneyre *et al.*, 2002; Polley *et al.*, 2003). The mechanisms that lead to higher responses of forb species than grasses to elevated CO₂ are not exactly known. However, results from our experiment indicate that mycorrhizae may significantly contribute to this differential response through altering nutrient acquisition of the coexisting plants. Compared with grasses, forb roots have a much smaller surface area per unit of root mass, potentially limiting their access to nutrients. CO₂-enhancement of mycorrhizae may, therefore, benefit forbs more than grasses in increasing root surface and nutrient interception, particularly of less mobile nutrients like P (Rouhier & Read, 1998; Syvertsen & Graham, 1999). Increased N and ¹⁵N acquisition of plantago (Figs 3 and 4; Table 2) and reduced N in the Test compartments (Fig. 5) in our experiment provide direct evidence illustrating that elevated CO₂ promoted mycorrhizally mediated N uptake by *P. lanceolata* but not by fescue plants. This mycorrhizal modification of N allocation among coexisting plants can be important in understanding plant N acquisition and plant interaction under future CO₂ scenarios (Johnson *et al.*, 2003; Hu *et al.*, 2005).

One may argue that root exclusion through the nylon mesh in our microcosms is artificial, which does not represent the true situation in field soil where roots of mycorrhizal and nonmycorrhizal hosts enmesh each other. However, mycorrhizal hyphae may allow their hosts to access N and other nutrients in some microsites where plant roots are unable to reach (Schimel & Bennett, 2004). CO₂-enhancement of mycorrhizae may be a major mechanism through which plants increase nutrient interception and transport under elevated CO₂ (Hu *et al.*, 2001, 2005). In addition, labile C inputs associated with mycorrhizal growth and turnover stimulate localized microbial activity and N mineralization (Hodge *et al.*, 2001). Because of the vicinity of mycorrhizal hyphae to those hotspots, their hosts have the advantage to access this newly released N source

(presumably other nutrients as well) over mycorrhizal-poor fescue plants.

It is very interesting to note that in our experiments, mycorrhizal root infection of both E+ and E- tall fescue varieties was extremely low and was not altered by elevated CO₂. Low mycorrhizal colonization of tall fescue roots has been attributed to the toxins produced by the endophyte symbionts (Chu-Chou *et al.*, 1992; Guo *et al.*, 1992; Müller, 2003). However, this assumption has not been vigorously tested. A recent study actually documented a positive interaction between *Neotyphodium* endophytes and arbuscular mycorrhizae in grass *Bromus setifolius* (Novas *et al.*, 2005). Although toxin production was not ascertained in our experiment, low mycorrhizal infection in E- plants suggests that endophytic toxins might not directly contribute to mycorrhizal inhibition, at least in the varieties examined in our experiments.

Because both endophytes and mycorrhizae depend on the C sources from the hosts, it has also been suggested that competition for C sources between those two fungal groups exist (Vicari *et al.*, 2002) and lower C availability in roots of endophytic plants might inhibit mycorrhizal infection and growth. Similarly, Vicari *et al.* (2002) showed that the beneficial effect of the foliar endophyte on perennial ryegrass, in terms of insect resistance, was reduced by mycorrhizae. When environmental stresses do not exist, endophytes might even behave as a C sink that reduces the competitiveness of host plants (Faeth & Fagan, 2002). If this hypothesis holds true, however, mycorrhizae would have taken advantage of enhanced C availability under elevated CO₂ or in the absence of endophyte in the host plants in our experiments. As the biomass remained statistically same for E+ and E- plants in Experiment 2, this 'C sink' hypothesis could not explain why the biomass of *P. lanceolata* was higher in the presence of E+ than E- plants. The alternative explanation is that E+ plants may increase resource availability for coexisting plants. Van Hecke *et al.* (2005) have recently shown significantly greater amount of soluble organic C and in turn higher microbial activity in the rhizosphere of E+ than E- tall fescue plants, enhancing nutrient mineralization and release from organic materials.

Responses of mycorrhizal and endophytic fungi to global change components may be important in understanding species interactions in many grasslands where mycorrhizal forbs and endophytic grasses coexist under future climatic conditions. On one hand, many grasses and forbs in grasslands are highly mycorrhizal (Smith *et al.*, 1999; Johnson *et al.*, 2003) and may confer additional competitive advantages over mycorrhizae-poor plants such as tall fescue under future CO₂ conditions, particularly in low nutrient soils. On the other hand, it

remains unclear how global change factors affect the activities of endophytic fungi. Our results suggest that endophytes may be less sensitive than mycorrhizae to elevated CO₂. Newman *et al.* (2003) observed that the presence of endophytes in tall fescue buffered their host response to elevated CO₂ by reducing the difference in the content of different crude proteins between ambient and elevated CO₂. Similarly, Hunt *et al.* (2005) have recently found that in the endophyte (*N. lolii*)-free ryegrass, soluble protein was significantly lower under elevated than ambient CO₂ and this difference was absent in endophyte-infected plants. In our experiments, plant biomass and N acquisition in *P. lanceolata* was less responsive to elevated CO₂ in the presence of endophytic tall fescue plants, although tall fescue plants were not affected (Fig. 7). In addition, it is well documented that endophytic fungi confer competitive advantage for host plants through enhancing their resistance to environmental stresses (Clay & Holah, 1999). Mineral N inputs have been shown to enhance toxin production by endophytic fungi in tall fescue (Belesky *et al.*, 1988; Arechavaleta *et al.*, 1992), but how the resulting changes influence mycorrhizal colonization and functioning remains unexplored. Together, these results indicate that the interactive effect of multiple global change components on mycorrhizae and foliar endophytes, and the resulting impact on species interactions warrant further investigation.

In summary, results obtained from our microcosm experiments showed that elevated CO₂ favored *P. lanceolata* over *F. arundinacea*, although the growth of both species was stimulated. Elevated CO₂ enhanced mycorrhizae in *P. lanceolata*, but did not have any significant effects on mycorrhizal colonization of *F. arundinacea* roots regardless of the status of endophytic infection. Neither the presence of endophytes nor mycorrhizal inoculation enhanced N acquisition and biomass accumulation in *F. arundinacea*. CO₂-enhancement of N acquisition in *P. lanceolata* through mycorrhizae likely predominates the differential effect of elevated CO₂ on two species. However, the role of endophytes in mediating the interactions between endophytic plants and their coexisting species under future CO₂ conditions remains unknown. Future studies are warranted to examine whether findings from this microcosm system hold true in field conditions and for other plant species and endophyte complexes under multiple environmental stresses.

Acknowledgements

We thank Howard Sanford for ¹⁵N measurements and Lisa Lentz for total C and N determinations at the Soil Analytical Lab, NCSU. Thanks were also extended to Nick Hill and Jim Green

for their constructive suggestions for experimental design, Jeff Barton for his technical help in maintaining the CSTR facility and to Betsy Randall-Schadel for her help in assessing the endophyte status in seeds and tillers of fescue plants. This study was supported by the Grants from the US Department of Agriculture (NRI-2000-00531) and the National Science Foundation (DEB-00-01686) to NC State University. Xin Chen was partially supported by a scholarship from the Pao Foundation Scholarship and a Grant from the National Science Foundation of China (NSFC, No. 30228005).

References

- Applebee TA, Gibson DJ, Newman JA (1999) Elevated atmospheric carbon dioxide alters the effects of allelochemicals production by tall fescue on alfalfa seedlings. *Transactions of the Illinois State Academy of Science*, **92**, 23–31.
- Arechavaleta M, Bacon CW, Plattner RD, Hoveland CS, Radcliff DE (1992) Accumulation of ergopeptide alkaloids in symbiotic tall fescue grown under deficits of soil-water and nitrogen-fertilizer. *Applied and Environmental Microbiology*, **58**, 857–861.
- Arnold AE, Mejia LC, Kylo D, Rojas EI, Maynard Z, Robbins N, Herre EA (2003) Fungal endophytes limit pathogen damage in a tropical tree. *Proceedings of National Academy of Sciences, USA*, **100**, 15649–15654.
- Bacon CW, Porter JK, Robbins JD, Luttrell ES (1977) *Epichloetypophina* from toxic tall fescue grasses. *Applied and Environmental Microbiology*, **34**, 576–581.
- Ball DM, Pedersen JA, Lacefield GD (1993) The tall-fescue endophyte. *American Scientist*, **81**, 370–379.
- Barton SB, Nestor RL (1979) Forage and weed species and grazing management systems on permanent pastures in the Allegheny Highlands of West Virginia. www.caf.wvu.edu/~forage/library/forglvt/bulletins/bul670.pdf
- Belesky DP, Stuedemann JA, Plattner RD, Wilkinson SR (1988) Ergopeptide alkaloids in grazed tall fescue. *Agronomy Journal*, **80**, 209–212.
- Booker FL, Shafer SR, Wei CM, Horton SJ (2000) Carbon dioxide enrichment and nitrogen fertilization effects on cotton (*Gossypium hirsutum* L.) plant residue chemistry and decomposition. *Plant and Soil*, **220**, 89–98.
- Bree JP (1994) *Acremonium* endophyte interactions with enhanced plant resistance to insects. *Annual Review of Entomology*, **39**, 401–423.
- Chu-Chou M, Guo B, An ZQ, Hendrix JW, Ferriss RS, Siegel MR, Dougherty CT, Burrus PB (1992) Suppression of mycorrhizal fungi in fescue by the *Acremonium-coenophialum* endophyte. *Soil Biology and Biochemistry*, **24**, 633–637.
- Clay K, Holah J (1999) Fungal endophyte symbiosis and plant diversity in successional fields. *Science*, **285**, 1742–1744.
- Clay K, Schardl C (2002) Evolutionary origins and ecological consequences of endophyte symbiosis with grasses. *American Naturalist*, **151**, 264–267.
- Elmi AA, West CP (1995) Endophyte infection effects on stomatal conductance, osmotic adjustment and drought recovery of tall fescue. *New Phytologist*, **131**, 61–67.
- Faeth SH, Fagan WF (2002) Fungal endophytes: common host plant symbionts but uncommon mutualists. *Integrative and Comparative Biology*, **42**, 360–368.

- Giovannetti M, Mosse B (1980) Evaluation of techniques for measuring vesicular arbuscular mycorrhizal infection in roots. *New Phytologist*, **84**, 489–500.
- Glenn AE, Bacon CW, Price R, Hanlin RT (1996) Molecular phylogeny of *Acremonium* and its taxonomic implications. *Mycologia*, **88**, 369–383.
- Guo BZ, Hendrix JW, An ZQ, Ferriss RS (1992) Role of *Acremonium* endophyte of fescue on inhibition of colonization and reproduction of mycorrhizal fungi. *Mycologia*, **84**, 882–885.
- Gwinn KD, Bernard EC (1993) Interactions of endophyte-infected grasses with nematodes *Meloidogyne marylandi* and *Pratylenchus scribneri*. In: *Proceedings of the Second International Conference on Acremonium/Grass Interactions* (ed. Hume DE), pp. 156–160. AgResearch, Palmerston North, New Zealand.
- Hartwig UA, Wittmann P, Raun RB *et al.* (2002) Arbuscular mycorrhiza infection enhances the growth response of *Lolium perenne* to elevated atmospheric CO₂. *Journal of Experimental Botany*, **53**, 1207–1213.
- Hiatt EE, Hill NS, Bouton JH, Stuedemann JA (1999) Tall fescue endophyte detection: commercial immunoblot test kit compared with microscopic analysis. *Crop Science*, **39**, 796–799.
- Hill NS, Thompson FN, Dawe DL, Stuedemann JA (1994) Antibody binding of circulating ergopeptide alkaloids in cattle grazing tall fescue. *American Journal of Veterinary Research*, **55**, 419–425.
- Hodge A, Campbell CD, Fitter AH (2001) An arbuscular mycorrhizal fungus accelerates decomposition and acquires nitrogen directly from organic material. *Nature*, **413**, 297–299.
- Hu S, Chapin FS III, Firestone MK, Field CB, Chiariello NR (2001) Nitrogen limitation of microbial decomposition in a grassland under elevated CO₂. *Nature*, **409**, 188–191.
- Hu SJ, Firestone MK, Chapin FS (1999) Soil microbial feedbacks to atmospheric CO₂ enrichment. *Trends in Ecology and Evolution*, **14**, 433–437.
- Hu S, Wu J, Burkey KO, Firestone MK (2005) Plant and microbial N acquisition under elevated atmospheric CO₂ in two microcosm experiments with annual grasses. *Global Change Biology*, **11**, 223–233.
- Hunt MG, Rasmussen S, Newton PCD, Parsons AJ, Newman JA (2005) Near-term impacts of elevated CO₂, nitrogen and fungal endophyte-infection on *Lolium perenne* L. growth, chemical composition and alkaloid production. *Plant, Cell and Environment*, **28**, 1345–1354.
- IPCC. (2001) *Climate Change 2001: The Scientific Basis*. Cambridge University Press, Cambridge.
- Johnson NC, Wolf J, Koch GW (2003) Interactions among mycorrhizae, atmospheric CO₂ and soil N impact plant community composition. *Ecology Letters*, **6**, 532–540.
- Latch GCM (1993) Physiological interactions of endophytic fungi and their host–biotic stress tolerance imparted to grasses by endophytes. *Agriculture, Ecosystems and Environment*, **44**, 143–156.
- Leadley PW, Niklaus PA, Stocker R, Korner C (1999) A field study of the effects of elevated CO₂ on plant biomass and community structure in a calcareous grassland. *Oecologia*, **118**, 39–49.
- Lyons PC, Evans JJ, Bacon CW (1990) Effects of the fungal endophyte *Acremonium coenophialum* on nitrogen accumulation and metabolism in tall fescue. *Plant Physiology*, **92**, 726–732.
- Marks S, Clay K (1990) Effects of CO₂ enrichment, nutrient addition, and fungal endophyte-infection on the growth of 2 grasses. *Oecologia*, **84**, 207–214.
- Marks S, Lincoln DE (1996) Antiherbivore defense mutualism under elevated carbon dioxide levels: a fungal endophyte and grass. *Environmental Entomology*, **25**, 618–623.
- Matthews JW, Clay K (2001) Influence of fungal endophyte infection on plant–soil feedback and community interactions. *Ecology*, **82**, 500–509.
- Müller J (2003) Artificial infection by endophytes affects growth and mycorrhizal colonization of *Lolium perenne*. *Functional Plant Biology*, **30**, 419–424.
- Müller CB, Krauss J (2005) Symbiosis between grasses and asexual fungal endophytes. *Current Opinion in Plant Biology*, **8**, 450–456.
- Newman JA, Abner ML, Dado RG, Gibson DJ, Brookings A, Parsons AJ (2003) Effects of elevated CO₂, nitrogen and fungal endophyte-infection on tall fescue: growth, photosynthesis, chemical composition and digestibility. *Global Change Biology*, **9**, 425–437.
- Novas MV, Cabral D, Godeas AM (2005) Interaction between grass endophytes and mycorrhizas in *Bromus setifolius* from Patagonia, Argentina. *Symbiosis*, **40**, 23–30.
- O'Connor PJ, Smith SE, Smith EA (2002) Arbuscular mycorrhizas influence plant diversity and community structure in a semi-arid herbland. *New Phytologist*, **154**, 209–218.
- Owensby CE, Ham JM, Knapp AK, Auen LM (1999) Biomass production and species composition change in a tallgrass prairie ecosystem after long-term exposure to elevated atmospheric CO₂. *Global Change Biology*, **5**, 497–436.
- Polley HW, Johnson HB, Derner JD (2003) Increasing CO₂ from subambient to superambient concentrations alters species composition and increases above-ground biomass in a C₃/C₄ grassland. *New Phytologist*, **160**, 319–327.
- Potvin C, Vasseur L (1997) Long-term CO₂ enrichment of pasture community: species richness, dominance, and succession. *Ecology*, **78**, 666–677.
- Richardson MD, Hoveland CS, Bacon CW (1993) Photosynthesis and stomatal conductance of symbiotic and nonsymbiotic tall fescue. *Crop Science*, **33**, 145–149.
- Rillig MC, Field CB, Allen MF (1999) Fungal root colonization responses in natural grasslands after long-term exposure to elevated atmospheric CO₂. *Global Change Biology*, **5**, 577–585.
- Rouhier H, Read DJ (1998) The role of mycorrhiza in determining the response of *Plantago lanceolata* to CO₂ enrichment. *New Phytologist*, **139**, 367–373.
- Rudgers JA, Koslow JM, Clay K (2004) Endophyte fungi alter relationships between diversity and ecosystem properties. *Ecology Letters*, **7**, 42–51.
- Saikkonen K (2000) Kentucky 31, far from home. *Science*, **287**, 1887.
- Schardl CL, Leuchtman A, Spiering MJ (2004) Symbioses of grasses with seedborne fungal endophytes. *Annual Review of Plant Biology*, **55**, 315–340.
- Schimel JP, Bennett J (2004) Nitrogen mineralization: challenges of a changing paradigm. *Ecology*, **85**, 591–602.

- Smith DM, Hartnett DC, Wilson GWT (1999) Interacting influence of mycorrhizal symbiosis and competition on plant diversity in tallgrass prairie. *Oecologia*, **121**, 574–582.
- Smith SE, Read DJ (1997) *Mycorrhizal Symbiosis*. Academic Press, San Diego, CA, USA.
- Staddon PL, Fitter AH (1998) Does elevated atmospheric carbon dioxide affect arbuscular mycorrhizas? *Trends in Ecology and Evolution*, **13**, 455–458.
- Staddon PL, Fitter AH, Graves JD (1999) Effect of elevated atmospheric CO₂ on mycorrhizal colonization, external mycorrhizal hyphal production and phosphorus inflow in *Plantago lanceolata* and *Trifolium repens* in association with the arbuscular mycorrhizal fungus *Glomus mosseae*. *Global Change Biology*, **5**, 347–358.
- Syvertsen JP, Graham JH (1999) Phosphorus supply and arbuscular mycorrhizas increase growth and net gas exchange responses of two *Citrus* spp. grown at elevated CO₂. *Plant and Soil*, **208**, 209–219.
- Teyssonneyre F, Picon-cochard C, Falcimagne R, Soussana JF (2002) Effects of elevated CO₂ and cutting frequency on plant community structure in a temperate grassland. *Global Change Biology*, **8**, 1034–1046.
- Treseder KK (2004) A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric CO₂ in field studies. *New Phytologist*, **164**, 347–355.
- Tu C, Booker FL, Watson DM, Chex X, Rufty TW, Shi W, Hu S (2006) Mycorrhizal mediation of plant N acquisition and residue decomposition: impact of mineral N inputs. *Global Change Biology*, **12**, 793–803.
- van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, Wiemken A, Sanders IR (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature*, **396**, 69–72.
- Van Hecke MM, Treonis AM, Kaufman JR (2005) How does the fungal endophyte *Neotyphodium coenophialum* affect tall fescue (*Festuca arundinacea*) rhizodeposition and soil microorganisms? *Plant and Soil*, **275**, 101–109.
- Vicari M, Hatcher PE, Ayres PG (2002) Combined effect of foliar and mycorrhizal endophytes on an insect herbivore. *Ecology*, **83**, 2452–2464.
- Zanetti S, Hartwig UA, Luscher A *et al.* (1996) Stimulation of symbiotic N₂ fixation in *Trifolium repens* L. under elevated atmospheric pCO₂ in a grassland ecosystem. *Plant Physiology*, **112**, 575–583.